1

Title: Prognostic value of programmed death ligand 1 and programmed death 1 expression in thymic

carcinoma

Authors: Shintaro Yokoyama, Hiroaki Miyoshi, Kazutaka Nakashima, Joji Shimono, Toshihiro

Hashiguchi, 1,2 Masahiro Mitsuoka, 1 Shinzo Takamori, 1 Yoshito Akagi, 1 and Koichi Ohshima 2

Author's affiliations: ¹Department of Surgery, Kurume University School of Medicine, Kurume,

Japan

²Department of Pathology, Kurume University School of Medicine, Kurume, Japan

Running title: PD-L1 and PD-1 expression in thymic carcinoma

Keywords: thymic carcinoma, thymic epithelial tumor, programmed death ligand 1, programmed

death 1, tumor immunology.

Conflicts of interest: There are no conflicts of interest to declare.

Corresponding author: Hiroaki Miyoshi, MD, PhD, Department of Pathology, Kurume University

School of Medicine, 67 Asahi-machi, Kurume, 8300011, Japan

Telephone: +81-942-31-7547. Fax: +81-942-35-8967.

E-mail: miyoshi_hiroaki@med.kurume-u.ac.jp

Word count: 3,687 words

Total number of figures and tables: 8 (including 2 supplementary tables)

STATEMENT OF TRANSLATIONAL RELEVANCE

Programmed death 1/programmed death ligand 1 (PD-1/PD-L1) pathway contributes to evasion of host immunity through negative regulation of T cells in the tumor microenvironment. Although the prognostic implications of PD-1/PD-L1 expression have been reported in various malignancies, its role in thymic carcinoma remains unclear. Herein, for the first time, we provide evidence that high PD-L1 expression in tumor cells is associated with favorable prognosis, whereas an increase in the number of PD-1⁺ tumor infiltrating lymphocytes is correlated with poor prognosis in thymic carcinoma. Significant proportional correlations were also observed between PD-L1 expression and cytotoxic T lymphocyte abundance. These indicators can be used to stratify patients for more effective clinical management. More importantly, our findings may also contribute to establishing a basis for the potential application of targeting the PD-1/PD-L1 pathway via immunotherapy as a novel treatment for thymic carcinoma.

ABSTRACT

Purpose: The immune checkpoint of the programmed death 1/programmed death ligand 1 (PD-1/PD-L1) pathway is believed to play an important role in evasion of host antitumor immune surveillance in various malignancies; however, little is known about its role in thymic carcinoma. This study investigated PD-1/PD-L1 expression and its association with clinicopathological features, the expression of immune-related proteins in tumor infiltrating lymphocytes (TILs), and patient prognosis.

Experimental Design: PD-L1 and PD-1 expression was evaluated by immunohistochemistry in 25 thymic carcinoma tissue specimens. Copy number alterations of the *PD-L1* gene in 11 cases were assessed in formalin-fixed, paraffin-embedded material using quantitative real-time PCR.

Results: Compared to normal subjects, 3 thymic carcinoma patients showed an increase in *PD-L1* copy number, whereas 8 did not. PD-L1 was significantly overexpressed in cases with copy number gain as compared to normal cases. High PD-L1 expression was associated with higher disease-free and overall survival rates as compared to cases with low expression. Prognostic analysis revealed low PD-L1 expression and high number of PD-1⁺ TILs as significant predictors of poor survival, together with Masaoka-Koga stage IVa/IVb disease and incomplete resection. In the quantitative analysis of TILs, PD-L1 expression correlated proportionally with the number of infiltrating cytotoxic T lymphocytes.

Conclusions: Here, for the first time, we report that PD-L1 and PD-1 expression might be useful

prognostic predictors in thymic carcinoma. Further studies are expected to substantiate the prognostic value of PD-L1 and PD-1 expression, and the potential efficacy of targeting the PD-1/PD-L1 pathway in thymic carcinoma via immunotherapy.

INTRODUCTION

Thymic epithelial tumors are the most common type of malignancy arising in the anterior mediastinum and include thymoma and thymic carcinoma; the latter is a relatively rare, high grade malignancy that directly invades adjacent structures and also undergoes distant metastasis. The most effective therapeutic modality for treating thymic carcinoma is surgical complete resection, ^{1, 2} which increases the 5-year overall survival from 33.3-65.0% to 53.0-65.7%. Moreover, thymic carcinoma is widely recognized to respond poorly to many chemotherapeutic agents, with an objective response rate of 20-36%, although concomitant administration of carboplatin and paclitaxel yields longer survival. ^{9, 10} Second-line treatments have also met with varying degrees of success, whereas therapeutic efficacy of sunitinib, an oral tyrosine kinase inhibitor, was recently reported in refractory patients previously treated with platinum-based chemotherapy. As such, there is a need for more effective therapeutic strategies, which requires detailed knowledge of underlying molecular oncogenic mechanisms.

Programmed death ligand 1 (PD-L1), which is expressed by antigen-presenting cells, is an immunomodulatory glycoprotein that acts as a negative regulator in immune responses. ¹² Programmed death 1 (PD-1) receptor is expressed on T cells and binds PD-L1 to inhibit phosphatidylinositol 3-kinase/Akt, which in turn suppresses T cell interleukin-2 production and reduces T cell proliferation and survival. ¹³ The PD-1/PD-L1 pathway has recently been shown to play pivotal roles in normal immune system regulation. ¹³ Furthermore, PD-L1 is also upregulated in

various malignancies and facilitates tumor progression by inhibiting T cell responses, allowing tumors to evade host immune surveillance mechanisms. ¹⁴ Indeed, PD-L1 expression and PD-1⁺ tumor infiltrating lymphocyte (TIL) abundance have been reported to be prognostic factors in various malignancies. ¹⁵⁻²¹ In previous reports on PD-L1 expression in thymic epithelial tumors, it was stated that high PD-L1 expression was associated with an advanced Masaoka-Koga stage and more aggressive histologies such as type B2/B3 thymoma or thymic carcinoma. ²²⁻²⁴ In contrast, survival analyses based on PD-L1 expression were inconsistent, as one study determined a significant association between high PD-L1 expression and poor overall survival (OS) in an age- and sex-adjusted analysis, whereas the others did not find any prognostic impact. Additionally, PD-1 expression in TILs has still not been examined in any detail. Finally, no study has evaluated PD-1/PD-L1 expression in thymic carcinoma as a subset of thymic epithelial tumors, in distinction from thymoma.

Blocking PD-1/PD-L1 signaling is a potential immunotherapeutic strategy for restoring antitumor T cell responses in various malignancies. Anti-PD-1 antibody has already been approved for clinical use in malignant melanoma, non-small cell lung cancer, and renal cell carcinoma, whereas anti-PD-L1 antibody has demonstrated therapeutic efficacy in patients resistant to conventional chemotherapeutic agents in clinical trials.²⁵ Immunotherapy targeting the PD-1/PD-L1 pathway is therefore expected to improve treatments of malignancies, particularly advanced, chemotherapy-resistant, or metastatic cases.

To this end, the present study investigated PD-L1 expression in thymic carcinoma and its correlation with clinicopathological features including PD-1⁺ TIL numbers and patient prognosis. Copy number alterations of the *PD-L1* gene were also examined, and their relationship to PD-L1 expression was assessed.

MATERIALS AND METHODS

Patients

Formalin-fixed, paraffin-embedded (FFPE) tissue samples acquired from 25 newly diagnosed consecutive thymic carcinoma patients at Kurume University Hospital between 1999 and 2015 were reviewed. All pathological diagnoses were made by two experienced pathologists (MH and OK) based on the 2015 World Health Organization classification with immunohistochemical detection of cluster of differentiation (CD)5 and CD117 as required. None of the samples included immature lymphocytes positive for terminal deoxynucleotidyl transferase, and relapsed cases or cases of pathologically combined thymoma and thymic carcinoma were excluded. Each patient had undergone clinical follow-up at least every 6 months after diagnosis. The samples and medical records used in this study were approved by the Research Ethics Committee of Kurume University and conformed to the ethical guidelines of the Declaration of Helsinki. Informed consent to use materials and clinical information was obtained from all patients.

Tissue microarray construction

Tissue microarrays (TMAs) were prepared using a standard tissue microarrayer (Azumaya Corp., Tokyo, Japan). A 3000-μm core was obtained from the central area of each tumor to include the maximum number of tumor cells. This procedure was based on small biopsy specimens, which are often clinically obtained using image-guided techniques, where the central area of the tumor are likely to be extracted. The TMA blocks were sectioned at a thickness of 2.5 μm and stained with hematoxylin and eosin or used for immunohistochemistry (IHC).

Immunohistochemistry

The primary antibodies used for IHC were as follows: rabbit monoclonal anti-PD-L1 (EPR1161[2]; Abcam, Cambridge, UK) at 1:200; mouse monoclonal anti-PD-1 (NAT105; Abcam) at 1:50; mouse monoclonal anti-CD56 (1B6; Leica Biosystems, Wetzlar, Germany) at 1:100; mouse monoclonal anti-T cell intracellular antigen 1 (TIA-1) (2G9A10F5; Beckman Coulter, Brea, CA) at 1:200; and mouse monoclonal anti-granzyme B (GrB-7; Merck Millipore, Billerica, MA) at 1:500. Tissue sections were deparaffinized and then rehydrated with H₂O. Antigen retrieval was performed by heating in a microwave oven at 95°C for 20 minutes in ethylenediaminetetraacetic acid buffer (pH 8.0). Endogenous peroxidase activity was blocked by incubating in 3% H₂O₂ for 5 minutes. TMA slides were incubated in primary antibodies as follows: 16 hours in a humidified chamber at 4°C for PD-L1 and 30 minutes at room temperature for PD-1/CD56/TIA-1/granzyme B. Samples were then incubated with horseradish peroxidase-conjugated anti-rabbit/mouse secondary antibody of the Dako REAL EnVision Detection System (Dako, Glostrup, Denmark) for 30 minutes at room temperature.

The immunoreaction was visualized by treatment with diaminobenzidine chromogen (Dako) for 5 minutes. Each slide was evaluated by two investigators (YS and MH) who were blinded to any clinical information, and discrepancies were debated until a consensus was reached.

Appraisal of PD-L1 expression

Quantification of PD-L1 expression

PD-L1 expression was assessed under 400-fold magnification using an optical microscope in an entire field for each TMA spot and was quantified by the H-score approach, in which the proportion and intensity of tumor cells positive for PD-L1 is determined.^{23,27} Specifically, the PD-L1 staining intensity of individual tumor cells was graded from 0 to 3 (0, negative; 1, weak; 2, moderate; 3, strong). The proportion of tumor cells exhibiting each intensity level with respect to the entire tumor cell population was calculated. The H-score was defined as the sum of the values obtained by multiplying the staining intensity and proportion, and ranged from 0 to 300 (Fig. 1).

PD-L1 expression (H-score) cutoff value determination

The optimal cutoff value for PD-L1 expression was defined based on the receiver operating characteristic curve and Youden's index. ^{28, 29} Here, Masaoka-Koga stage and surgical curability were taken into account as variables that significantly affect prognosis. ^{8, 30} Ultimately, either Masaoka-Koga stage I/IIa/IIb/III disease who all performed complete resection or stage IVa/IVb who undergone diagnostic biopsy was applied as a dichotomous variable, with PD-L1 expression as a continuous variable. Youden's index revealed as optimal cutoff value an H-score of 20; cases with

PD-L1 expression higher than 20 and equal to or lower than 20 were defined as cases with high and low PD-L1 expression, respectively.

Immunohistochemical analysis of TILs expressing PD-1, TIA-1, and granzyme B

The numbers of PD-1⁺, TIA-1⁺, and granzyme B⁺ TILs were respectively enumerated in the area showing the strongest staining for each antibody. In this quantification, because PD-1⁺ and granzyme B⁺ TILs were not infiltrated in tumor nests or around vessels penetrating the tumor, they were counted in the stroma among tumor nests. In contrast, TIA-1⁺ TILs were counted mainly in tumor nests or beside glandular structures within the tumor. However, in some cases they were counted in the stroma because of the absence of TIA-1⁺ TILs in these sites. All TILs were counted in 5 high-power fields (HPFs) under 400-fold magnification. The mean number of TILs that demonstrated positive staining for each antibody per HPF was calculated.

Quantitative real-time polymerase chain reaction analysis of PD-L1 copy number

Copy number analyses of *PD-L1* in 11 FFPE tissue samples were performed by quantitative real-time polymerase chain reaction (PCR). Samples with adequate tumor regions were dissected and total DNA was extracted using a QIAamp DNA Micro Kit (QIAGEN, Hilden, Germany). *PD-L1* copy number was evaluated using the TaqMan DNA Copy Number Assay (Hs03704252_cn; Applied Biosystems, Waltham, MA) and Taqman Copy Number Reference Assay *RNase P* (Applied Biosystems). Quantitative real-time PCR was performed using Taqman Universal Genotyping Master Mix (Applied Biosystems) according to the manufacturer's protocol on an Applied Biosystems 7500

real-time PCR system. Results were analyzed using CopyCaller Software (Applied Biosystems).

Statistical analysis

The relationship between PD-L1 positivity and clinicopathological features was evaluated using Fisher's exact test for categorical variables. The Mann-Whitney U test was used to compare the median value of continuous variables according to PD-L1 positivity as well as PD-L1 expression based on PD-L1 copy number status. Pearson's product-moment correlation coefficient was used to determine the correlation between PD-L1 expression and the number of TIA-1⁺ or granzyme B⁺ TILs. In the analysis of recurrence and survival, the starting point was defined as the day of surgical resection or biopsy. The end of the disease-free survival (DFS) period was defined as the day of recurrence, and that of the OS period was defined as the day of death or the date of last follow-up, respectively. Survival curves to estimate DFS and OS were determined by the Kaplan-Meier method, with comparison of each curve using a log-rank test. Cox proportional hazards regression models were used to assess the prognostic values of each factor. P-values declared in this study were all based on two-sided tests, and those less than 0.05 were interpreted as statistically significant. JMP version 11 software (SAS Institute Inc., Cary, NC) was used to perform all statistical analyses.

RESULTS

Clinicopathological findings

The baseline clinicopathological features of patients are summarized in Supplementary

Table S1. The median follow-up period was 33 months (range, 1-124). The 25 cases of thymic carcinoma included 17 males (68%) and 8 females (32%), with a median age of 63 years (range, 32-81). Surgical complete resection was achieved in 16 cases (64%), which was attained in all cases representing Masaoka-Koga stage I/IIa/IIb/III disease with microscopically confirmed negative surgical margin. In 9 cases (36%) of Masaoka-Koga stage IVa/IVb disease, only diagnostic biopsies were performed and specific nonsurgical treatments were administered. Four cases (25%) received neoadjuvant therapies, including 2 cases of radiotherapy (13%) and 2 of chemoradiotherapy (13%); subsequent surgical complete resections were successfully performed for all cases.

Immunohistochemical analysis of PD-L1 expression

PD-L1 was mainly expressed on the cell membrane with or without cytoplasm of tumor cells, exhibiting a near-uniform pattern in tumor regions (Fig. 1). The median value of PD-L1 expression was 100, with a range of 0-300 (95% confidence interval [CI], 77.8-154.4). According to our predefined cutoff value, 20/25 cases (80%) were classified as having high PD-L1 expression.

PD-L1 copy number analysis and association with PD-L1 expression

The validity of the *PD-L1* copy number assay was confirmed using 5 normal control cases of reactive tonsils, which had a mean copy number of 2.07 (95% CI, 1.83-2.31). The mean copy number in each case was determined by triplicate PCRs with adjustment by control cases, which revealed *PD-L1* gene copy number gains in 3 cases (27%), while 8 cases (73%) showed a normal *PD-L1* status (Fig. 2A). Cases with *PD-L1* copy number gain showed significantly higher PD-L1

expression than normal cases (P = 0.040, Fig. 2B).

Relationship between PD-L1 positivity and clinicopathological features

Table 1 depicts the distributions of clinicopathological characteristics according to positivity for PD-L1. PD-L1 expression status was not associated with age (P = 0.586), sex (P = 1.000), Masaoka-Koga stage (P = 0.312), histology (P = 1.000), tumor size (P = 0.838), or surgical curability (P = 0.312). Of note, the number of PD-1⁺ TILs was also not associated with PD-L1 expression status (P = 0.837).

Clinical outcome according to PD-L1 expression

On Kaplan-Meier analysis, OS of all patients was significantly longer in cases with high as compared to low PD-L1 expression (P = 0.010, Fig. 3A). Furthermore, significant associations were also encountered in the subgroup of patients with completely resected tumor (P = 0.0009, Fig. 3B) and the subgroup of patients with thymic squamous cell carcinomas (P = 0.005, Fig. 3C). Patients with high PD-L1 expression also had significantly better DFS than those with low PD-L1 expression (P = 0.004, Fig. 3D).

Potential prognostic factors affecting overall survival

Each clinicopathological factor, including PD-L1 and PD-1 expression status, was evaluated for its prognostic value (Table 2). In all patients, Masaoka-Koga stage IVa/IVb disease (hazard ratio [HR], 6.578; P = 0.004), incomplete resection (HR, 6.578; P = 0.004), low PD-L1 expression (HR, 5.837; P = 0.025), and increased number of PD-1⁺ TILs (HR, 1.496; P = 0.037) were identified as

significant predictors of poor survival in the univariate analysis. Furthermore, the statistical significance of both PD-L1 expression and PD-1⁺ TILs as prognostic factors was observed even in the squamous cell carcinoma subset, in which the HR for low PD-L1 expression was even higher (HR, 8.073; P = 0.014) than that in the entire cohort of thymic carcinoma patients.

Multivariate analysis of prognostic factors affecting overall survival

Although a larger multivariate model was used to identify prognostic factors, the results had limited value because of the small number of samples (Supplementary Table S2). For reference, all potential prognostic factors identified in the univariate analysis retained their significance in the multivariate analysis, represented as Masaoka-Koga stage (HR, 7.000; P = 0.005), surgical curability (HR, 7.000; P = 0.005), PD-L1 expression (HR, 6.854; P = 0.028), and the number of PD-1⁺ TILs (HR, 1.546; P = 0.026). In addition, these factors were prognostically significant in patients with squamous cell carcinomas.

Correlation between PD-L1 expression and cytotoxic T lymphocyte abundance

To further characterize TILs and assess the association between high PD-L1 expression and improved survival, we evaluated TIA-1 expression in TILs which is a representative marker of cytotoxic cells. Based on morphological findings, the acquired knowledge of antitumor immunity principled by T cells, ¹⁴ and the absence of CD56⁺ lymphocytes confirmed by our immunohistochemical surveillance, the TIA-1⁺ cells were validated to be cytotoxic T lymphocytes (CTLs). CTLs were subsequently counted in each case and statistical analysis represented that

PD-L1 expression was significantly correlated with the number of CTLs (r = 0.566, P = 0.003, Fig. 4). Additionally, the number of granzyme B⁺ lymphocytes was also investigated to determine the correlation between PD-L1 expression and the number of activated CTLs. However, there was little correlation between PD-L1 expression and activated CTL abundance (r = 0.051, P = 0.808).

DISCUSSION

This study demonstrated for the first time that high PD-L1 expression is associated with favorable OS in thymic carcinoma. This result was supported by the finding that high PD-L1 expression was correlated with an increase in CTL infiltration. Furthermore, we also found that an increase in the number of PD-1⁺ TILs predicted poor patient survival. These results suggest that the PD-1/PD-L1 pathway is a potential immunotherapeutic target in thymic carcinoma.

The *PD-L1* copy number analysis demonstrated that PD-L1 expression was significantly higher in cases with *PD-L1* copy number gain than in normal cases, implying that *PD-L1* gene alteration is an important mechanism of PD-L1 overexpression. Previous studies that focused on B cell lymphomas have also reported that *PD-L1* transcripts are more highly expressed in tissue samples and cell lines with increased *PD-L1* copy number than in normal cases, although it was not possible to make a direct comparison. To clarify the exact biological mechanisms underlying PD-L1 overexpression in thymic carcinoma, additional studies and data on transcriptional levels are required. Furthermore, *PD-L1* was found to be amplified in 27% of thymic carcinomas. Although

few studies have evaluated genetic copy number aberrations in thymic carcinoma, relationships between poor OS and *BCL2* copy number gain or *CDKN2A/B* copy number loss have been observed, the frequency of which was 42% and 14%, respectively.³³ Further studies on genetic alterations in the tumor are necessary to improve therapeutic strategies and survival for thymic carcinoma patients.

Thus far, few studies have examined PD-L1 expression in thymic epithelial tumors. 22-24 Confined to thymic carcinoma as a subset of thymic epithelial tumors, only one of these reports mentioned the prognostic role of PD-L1, finding that there was no difference in OS according to PD-L1 positivity.²³ However, our results indicated a significant association between high PD-L1 expression and better prognosis. This discrepancy may be attributable to differences in patient selection; the previous study evaluated thymoma and thymic carcinoma collectively, whereas thymomas and heterogeneous cases of combined thymoma and thymic carcinoma were excluded from the present study. We consider that thymoma and thymic carcinoma should be analyzed individually because they are widely known to vary in terms of prognosis and molecular biological profiles.²⁶ In fact, when the 25 thymic carcinoma samples evaluated here were analyzed along with 82 thymoma samples in our previous study,²⁴ OS was comparable between cases with high and low PD-L1 expression in both individual histology types; thymic carcinoma, however, demonstrated slightly higher PD-L1 positivity than thymoma, consistent with the previous report. Nevertheless, our study analyzed thymic carcinoma independently and found that high PD-L1 expression was associated with favorable survival. Thus, thymic carcinoma and thymoma should be investigated

independently to clarify the role of PD-L1 expression in each tumor type.

The prognostic value of PD-L1 expression has been investigated for various malignancies, including malignant melanoma, non-small cell lung cancer, and renal cell carcinoma. 34-36 Most studies have emphasized an association between high PD-L1 expression and poor survival based on the ability of PD-L1 to negatively regulate antitumor T cell responses. In contrast, high PD-L1 expression has been linked to better prognosis in some malignancies. 17, 35, 37 In these reports, a significant association between high PD-L1 expression and increased TILs was identified. 17, 34, 35, 37 Moreover, it was also shown that expression of the immunosuppressive molecules PD-L1, indoleamine-2, 3-dioxygenase, and FoxP3 by tumor cells was induced following CD8⁺ T cell infiltration or interferon-gamma production in murine in vivo models of melanoma.³⁸ This suggests that PD-L1 expression by tumor cells is upregulated as a result of a feedback mechanism against activation of host antitumor immunity. In addition, increases in TIA-1⁺ TILs have been reported as a favorable prognostic factor in some malignancies, although the activated status of CTLs was not taken into account. 39-41 Based on these considerations, the association between high PD-L1 expression and improved prognosis in thymic carcinoma may be explained by high PD-L1 expression following the activation of host antitumor immune responses initiated by infiltrated CTLs. In order to substantiate the determined prognostic value of PD-L1 expression in thymic carcinoma, investigation of the interactions between immune-related molecules including PD-L1 within the tumor microenvironment is required.

This is the first report to investigate PD-1⁺ TILs in thymic carcinoma, demonstrating that an increased number of these cells predicts poor patient prognosis. Similarly, abundant PD-1⁺ TILs were previously reported to be a poor prognostic factor in some malignancies. ^{19, 20} These outcomes may be attributed to activation of T cell-mediated antitumor immune suppression via PD-1/PD-L1 signaling. Some studies have reported a proportional correlation between PD-L1 expression in tumor cells and the number of PD-1⁺ TILs, which may reflect PD-1/PD-L1 pathway activation; however, others have found an inverse correlation. ^{20, 42, 43} Although we could not provide definitive considerations in this argument because of the inability to directly compare studies, the ability to predict patient survival based on the number of PD-1⁺ TILs may be useful for stratifying patients in clinical settings.

We found that PD-L1 was frequently overexpressed by tumor cells and PD-1⁺ TILs infiltrated in thymic carcinoma. In recent years, PD-1/PD-L1-targeted immunotherapy has been shown to be clinically effective and thus represents a promising therapeutic alternative in selected patients with malignant tumors. Such efficacy is also expected for thymic carcinoma, which responds poorly to various chemotherapeutic agents; however, this has not been examined yet. In some other malignancies, immunotherapy using PD-L1- and PD-1-based antibodies appeared to show enhanced activity, favoring PD-L1-positive tumors over PD-L1-negative tumors.^{25,44} Although the practical potency is still unknown, these findings support our statement that the clinical use of immunotherapeutic antibodies targeting the PD-1/PD-L1 pathway may be effective for treating thymic carcinoma based on high PD-L1 positivity.

It is still controversial which anti-PD-L1 antibody is the best to determine PD-L1 expression by IHC in tumor tissue samples. This is a critical issue because immunohistochemical PD-L1 positivity has been reported to predict therapeutic efficacy in anti-PD-1/PD-L1 antibody immunotherapy in several clinical trials, as mentioned previously. In the present study, the PD-L1 clone EPR1161[2], which recognizes the C-terminus extracellular domains of PD-L1, was employed. By contrast, previous reports studying PD-L1 in thymic epithelial tumors used clone 5H1, which targets extracellular domains of PD-L1, or E1L3N, which targets intracellular domains, respectively.^{22, 23} These three studies (including ours) demonstrated slightly different populations of cases with high PD-L1 expression in thymic carcinoma (58.7-80%), which may have resulted from the use of distinct antibodies and PD-L1 cutoff values. Although the specificity of each antibody in IHC has certainly been confirmed, it remains unknown whether or not a particular antibody can predict the therapeutic efficacy of anti-PD-1/PD-L1-antibody immunotherapy. To establish a valid eligibility criterion for the clinical use of these agents, further studies evaluating the optimal methods (including the determination of the anti-PD-L1 antibody used for IHC and measures other than immunohistochemical PD-L1 expression) to accurately predict therapeutic efficacy are necessary in these treatments.

There are some limitations to our results. First, although a certain level of significance was observed in the multivariate analysis, this study included a small sample size, and the statistical power may have been insufficient. Additional studies with larger samples are necessary to confirm

the prognostic value of PD-L1 and PD-1 in thymic carcinoma. Second, our results were based on a retrospective analysis, which may have resulted in selection bias because of the various therapeutic protocols that were used. Therefore, prospective studies are also needed to validate our findings.

In conclusion, the present study demonstrated that high PD-L1 expression by tumor cells was associated with favorable prognosis, whereas an increased number of PD-1⁺ TILs was correlated with poor prognosis in thymic carcinoma. These indicators can be used to stratify patients for more effective clinical management. Importantly, our findings provide a basis for the use of immunotherapy targeting the PD-1/PD-L1 pathway as a potential novel treatment in thymic carcinoma.

Acknowledgements

The authors thank Mayumi Miura, Kanoko Miyazaki, Yuki Morotomi, Chie Kuroki, and Kaoruko Nagatomo for their technical assistance, and Masaki Kashihara, Tatsuya Nishi, Daigo Murakami, and Ryouichi Matsumoto for their support in managing clinical samples. Critical discussions and advice from Masao Seto, Yasuo Sugita, Daisuke Niino, Fumiko Arakawa, Shinji Nakashima, Junichi Kiyasu, Hiroko Muta, Toshiaki Haraguchi, Naohiro Yoshida, Noriaki Yoshida, Daisuke Kurita, Yuya Sasaki, Keisuke Kawamoto, Kotaro Matsuda, Junko Miyoshi, Satoru Komaki, Reiji Muto, and Eriko Yanagida are also gratefully acknowledged.

References

- Kondo K, Monden Y. Therapy for thymic epithelial tumors: a clinical study of 1,320 patients from Japan. Ann Thorac Surg. 2003;76:878-84.
- Venuta F, Anile M, Diso D, Vitolo D, Rendina EA, De Giacomo T, et al. Thymoma and thymic carcinoma. Eur J Cardiothorac Surg. 2010;37:13-25.
- Suster S, Rosai J. Thymic carcinoma. A clinicopathologic study of 60 cases. Cancer. 1991;67:1025-32.
- 4. Ogawa K, Toita T, Uno T, Fuwa N, Kakinohana Y, Kamata M, et al. Treatment and prognosis of thymic carcinoma: a retrospective analysis of 40 cases. Cancer. 2002;94:3115-9.
- Yano M, Sasaki H, Yokoyama T, Yukiue H, Kawano O, Suzuki S, et al. Thymic carcinoma: 30 cases at a single institution. J Thorac Oncol. 2008;3:265-9.
- Moon JW, Lee KS, Shin MH, Kim S, Woo SY, Lee G, et al. Thymic epithelial tumors: prognostic determinants among clinical, histopathologic, and computed tomography findings.
 Ann Thorac Surg. 2015;99:462-70.
- Huang J, Rizk NP, Travis WD, Riely GJ, Park BJ, Bains MS, et al. Comparison of patterns of relapse in thymic carcinoma and thymoma. J Thorac Cardiovasc Surg. 2009;138:26-31.
- Weissferdt A, Moran CA. Thymic carcinoma, part 1: a clinicopathologic and immunohistochemical study of 65 cases. Am J Clin Pathol. 2012;138:103-14.
- 9. Schmitt J, Loehrer PJ, Sr. The role of chemotherapy in advanced thymoma. J Thorac Oncol.

- 2010;5:S357-60.
- Lemma GL, Lee JW, Aisner SC, Langer CJ, Tester WJ, Johnson DH, et al. Phase II study of carboplatin and paclitaxel in advanced thymoma and thymic carcinoma. J Clin Oncol. 2011;29:2060-5.
- 11. Thomas A, Rajan A, Berman A, Tomita Y, Brzezniak C, Lee MJ, et al. Sunitinib in patients with chemotherapy-refractory thymoma and thymic carcinoma: an open-label phase 2 trial. Lancet Oncol. 2015;16:177-86.
- Brown JA, Dorfman DM, Ma FR, Sullivan EL, Munoz O, Wood CR, et al. Blockade of programmed death-1 ligands on dendritic cells enhances T cell activation and cytokine production. J Immunol. 2003;170:1257-66.
- 13. Pedoeem A, Azoulay-Alfaguter I, Strazza M, Silverman GJ, Mor A. Programmed death-1 pathway in cancer and autoimmunity. Clin Immunol. 2014;153:145-52.
- 14. Dong H, Strome SE, Salomao DR, Tamura H, Hirano F, Flies DB, et al. Tumor-associated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. Nat Med. 2002;8:793-800.
- 15. Wu C, Zhu Y, Jiang J, Zhao J, Zhang XG, Xu N. Immunohistochemical localization of programmed death-1 ligand-1 (PD-L1) in gastric carcinoma and its clinical significance. Acta Histochem. 2006;108:19-24.
- 16. Nomi T, Sho M, Akahori T, Hamada K, Kubo A, Kanehiro H, et al. Clinical significance and

- therapeutic potential of the programmed death-1 ligand/programmed death-1 pathway in human pancreatic cancer. Clin Cancer Res. 2007;13:2151-7.
- 17. Droeser RA, Hirt C, Viehl CT, Frey DM, Nebiker C, Huber X, et al. Clinical impact of programmed cell death ligand 1 expression in colorectal cancer. Eur J Cancer. 2013;49:2233-42.
- 18. Ohigashi Y, Sho M, Yamada Y, Tsurui Y, Hamada K, Ikeda N, et al. Clinical significance of programmed death-1 ligand-1 and programmed death-1 ligand-2 expression in human esophageal cancer. Clin Cancer Res. 2005;11:2947-53.
- 19. Muenst S, Soysal SD, Gao F, Obermann EC, Oertli D, Gillanders WE. The presence of programmed death 1 (PD-1)-positive tumor-infiltrating lymphocytes is associated with poor prognosis in human breast cancer. Breast Cancer Res Treat. 2013;139:667-76.
- 20. Thompson RH, Dong H, Lohse CM, Leibovich BC, Blute ML, Cheville JC, et al. PD-1 is expressed by tumor-infiltrating immune cells and is associated with poor outcome for patients with renal cell carcinoma. Clin Cancer Res. 2007;13:1757-61.
- 21. Wahlin BE, Aggarwal M, Montes-Moreno S, Gonzalez LF, Roncador G, Sanchez-Verde L, et al. A unifying microenvironment model in follicular lymphoma: outcome is predicted by programmed death-1--positive, regulatory, cytotoxic, and helper T cells and macrophages. Clin Cancer Res. 2010;16:637-50.
- 22. Padda SK, Riess JW, Schwartz EJ, Tian L, Kohrt HE, Neal JW, et al. Diffuse high intensity PD-L1 staining in thymic epithelial tumors. J Thorac Oncol. 2015;10:500-8.

- 23. Katsuya Y, Fujita Y, Horinouchi H, Ohe Y, Watanabe S, Tsuta K. Immunohistochemical status of PD-L1 in thymoma and thymic carcinoma. Lung Cancer. 2015;88:154-9.
- 24. Yokoyama S, Miyoshi H, Nishi T, Hashiguchi T, Mitsuoka M, Takamori S, et al.
 Clinicopathologic and prognostic implications of programmed death ligand 1 expression in thymoma. Ann Thorac Surg. 2016;101:1361-9.
- Philips GK, Atkins M. Therapeutic uses of anti-PD-1 and anti-PD-L1 antibodies. Int Immunol.
 2015;27:39-46.
- 26. Travis WD, Brambilla E, Burke AP, Marx A, Nicholson AG. WHO classification of tumours of lung, pleura, thymus and heart (4th edition). Lyon, France: IARC; 2015;p.183-243.
- 27. McCarty KS, Jr., Miller LS, Cox EB, Konrath J, McCarty KS, Sr. Estrogen receptor analyses.
 Correlation of biochemical and immunohistochemical methods using monoclonal antireceptor antibodies. Arch Pathol Lab Med. 1985;109:716-21.
- 28. Akobeng AK. Understanding diagnostic tests 3: Receiver operating characteristic curves. Acta Paediatr. 2007;96:644-7.
- 29. Muenst S, Hoeller S, Dirnhofer S, Tzankov A. Increased programmed death-1⁺ tumor-infiltrating lymphocytes in classical Hodgkin lymphoma substantiate reduced overall survival. Hum Pathol. 2009;40:1715-22.
- 30. Hishida T, Nomura S, Yano M, Asamura H, Yamashita M, Ohde Y, et al. Long-term outcome and prognostic factors of surgically treated thymic carcinoma: results of 306 cases from a Japanese

- Nationwide Database Study. Eur J Cardiothorac Surg. 2016;49:835-41.
- 31. Twa DD, Chan FC, Ben-Neriah S, Woolcock BW, Mottok A, Tan KL, et al. Genomic rearrangements involving programmed death ligands are recurrent in primary mediastinal large B-cell lymphoma. Blood. 2014;123:2062-5.
- 32. Green MR, Monti S, Rodig SJ, Juszczynski P, Currie T, O'Donnell E, et al. Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. Blood. 2010;116:3268-77.
- 33. Petrini I, Meltzer PS, Zucali PA, Luo J, Lee C, Santoro A, et al. Copy number aberrations of *BCL2* and *CDKN2A/B* identified by array-CGH in thymic epithelial tumors. Cell Death Dis. 2012;3:e351.
- 34. Taube JM, Anders RA, Young GD, Xu H, Sharma R, McMiller TL, et al. Colocalization of inflammatory response with B7-H1 expression in human melanocytic lesions supports an adaptive resistance mechanism of immune escape. Sci Transl Med. 2012;4:127ra37.
- 35. Schmidt LH, Kummel A, Gorlich D, Mohr M, Brockling S, Mikesch JH, et al. PD-1 and PD-L1 expression in NSCLC indicate a favorable prognosis in defined subgroups. PLoS One. 2015;10:e0136023.
- 36. Thompson RH, Gillett MD, Cheville JC, Lohse CM, Dong H, Webster WS, et al. Costimulatory B7-H1 in renal cell carcinoma patients: Indicator of tumor aggressiveness and potential

- therapeutic target. Proc Natl Acad Sci U S A. 2004;101:17174-9.
- 37. Schalper KA, Velcheti V, Carvajal D, Wimberly H, Brown J, Pusztai L, et al. In situ tumor PD-L1 mRNA expression is associated with increased TILs and better outcome in breast carcinomas. Clin Cancer Res. 2014;20:2773-82.
- 38. Spranger S, Spaapen RM, Zha Y, Williams J, Meng Y, Ha TT, et al. Up-regulation of PD-L1, IDO, and T_{regs} in the melanoma tumor microenvironment is driven by CD8⁺ T cells. Sci Transl Med. 2013;5:200ra116.
- 39. Townsend KN, Spowart JE, Huwait H, Eshragh S, West NR, Elrick MA, et al. Markers of T cell infiltration and function associate with favorable outcome in vascularized high-grade serous ovarian carcinoma. PLoS One. 2013;8:e82406.
- 40. Koelzer VH, Dawson H, Andersson E, Karamitopoulou E, Masucci GV, Lugli A, et al. Active immunosurveillance in the tumor microenvironment of colorectal cancer is associated with low frequency tumor budding and improved outcome. Transl Res. 2015;166:207-17.
- 41. Zlobec I, Karamitopoulou E, Terracciano L, Piscuoglio S, Iezzi G, Muraro MG, et al. TIA-1 cytotoxic granule-associated RNA binding protein improves the prognostic performance of CD8 in mismatch repair-proficient colorectal cancer. PLoS One. 2010;5:e14282.
- 42. Taube JM, Klein A, Brahmer JR, Xu H, Pan X, Kim JH, et al. Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy.

 Clin Cancer Res. 2014;20:5064-74.

- 43. Kiyasu J, Miyoshi H, Hirata A, Arakawa F, Ichikawa A, Niino D, et al. Expression of programmed cell death ligand 1 is associated with poor overall survival in patients with diffuse large B-cell lymphoma. Blood. 2015;126:2193-201.
- 44. Meng X, Huang Z, Teng F, Xing L, Yu J. Predictive biomarkers in PD-1/PD-L1 checkpoint blockade immunotherapy. Cancer Treat Rev. 2015;41:868-76.

Figure legends

Figure 1. Immunohistochemical analysis of PD-L1 expression in thymic carcinoma. Hematoxylin and eosin staining (upper row, original magnification $40\times$) and PD-L1 immunostaining (lower row, original magnification $40\times$ and lower row inset, original magnification $200\times$) are shown. PD-L1 expression (H-score) in each case represents (A,D) 300 (proportions of each intensity score: 0 = 0%; 1 = 0%; 2 = 0%; 3 = 100%), (B,E) 120 (0 = 30%; 1 = 30%; 2 = 30%; 3 = 10%), and (C,F) 3 (0 = 97%; 1 = 3%; 2 = 0%; 3 = 0%)

Figure 2. *PD-L1* copy number status and PD-L1 expression in thymic carcinoma. A, quantitative real-time PCR-based *PD-L1* copy number analysis in 11 thymic carcinoma tissue samples. Each bar represents the results showing the mean (± SD) of triplicate PCRs. Cases 9, 10, and 11 showed copy number gain (gray bar). B, cases with copy number gain showed significantly higher PD-L1 expression than normal cases.

Figure 3. Kaplan-Meier survival curves based on PD-L1 expression in thymic carcinoma (black and orange lines, high and low PD-L1 expression, respectively). A, survival curves in all patients. B and C, survival curves in the subgroup of patients who had undergone complete resection (B) and those with squamous cell carcinoma (C). D, survival curves for DFS in patients who had undergone complete resection. In all analyses, cases with high PD-L1 expression experienced significantly better OS or DFS than cases with low PD-L1 expression.

Figure 4. Correlation between PD-L1 expression and the number of infiltrated CTLs. PD-L1

expression was significantly correlated with CTL abundance.

Table 1. Comparison between PD-L1 expression status and clinicopathological features

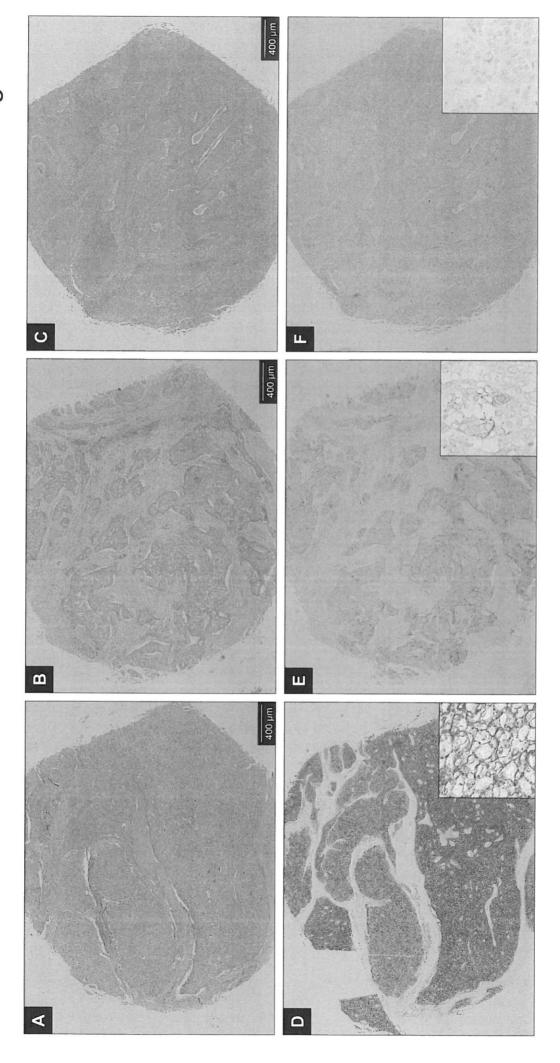
Characteristic	PD-L1 high	PD-L1 low		
Characteristic	n = 20 (80%)	n = 5 (20%)	P	
Age, years old (median, range)	66.5 [32-74]	62 [49-81]	0.586	
Sex, male	13 (65%)	4 (80%)	1.000	
Masaoka-Koga stage, IVa/IVb	6 (30%)	3 (60%)	0.312	
Histological type, SCC	15 (75%)	4 (80%)	1.000	
Tumor size, mm (median, range)	55.5 [13-90]	60 [25-80]	0.838	
Curability, complete resection	14 (70%)	2 (40%)	0.312	
PD-1 positive TILs / HPF (median, range)	1.5 [0-81.4]	1.6 [0-35.4]	0.837	

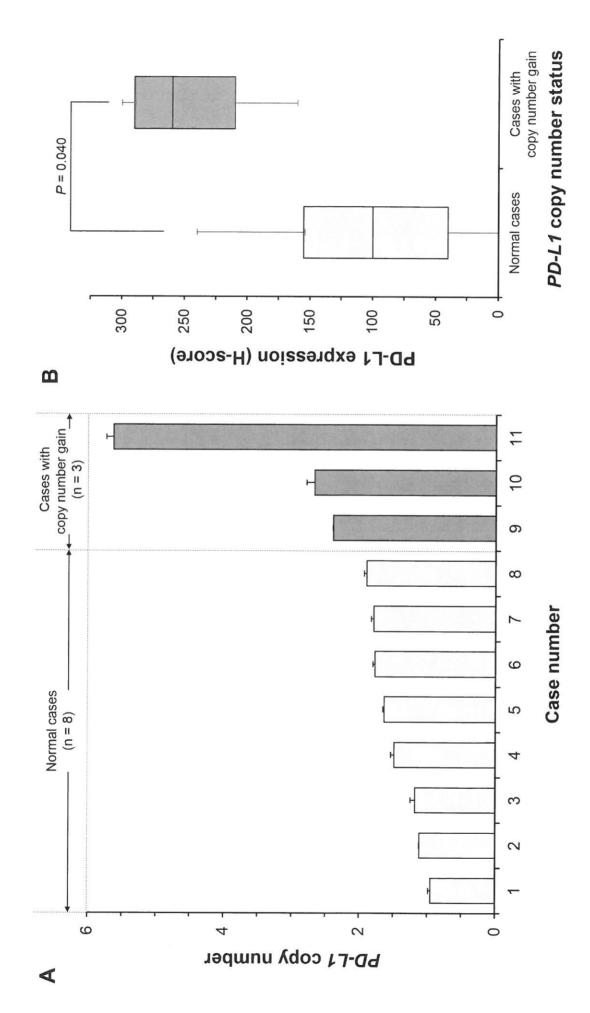
Abbreviations: HPF, high power field; PD-1, programmed death 1; PD-L1, programmed death ligand 1; SCC, squamous cell carcinoma; TIL, tumor infiltrating lymphocyte.

Table 2. Univariate analysis for prognostic factors affecting overall survival

Characteristic, risk factor	All cases (n = 25)			SCC (n = 19)		
	HR	[95% CI]	P	HR	[95% CI]	P
Age, < 63 years old	1.359	[0.408-5.217]	0.622	1.311	[0.370-5.175]	0.675
Sex, female	1.067	[0.279-3.541]	0.918	1.024	[0.260-3.612]	0.971
Masaoka-Koga stage, IVa/IVb	6.578	[1.789-30.971]	0.004	5.014	[1.304-24.008]	0.019
Histological type, SCC	3.195	[0.592-59.531]	0.205	-		-
Tumor size, > 56 mm	1.265	[0.359-4.279]	0.704	1.219	[0.341-4.844]	0.762
Curability, incomplete resection	6.578	[1.789-30.971]	0.004	5.014	[1.304-24.008]	0.019
PD-L1 expression, low expression	5.837	[1.265-30.167]	0.025	8.073	[1.538-59.367]	0.014
PD-1 positive TILs, per increase by 10 cells / HPF	1.496	[1.029-2.162]	0.037	1.476	[1.019-2.188]	0.041

Abbreviations: CI, confidence interval; HPF, high power field; HR, hazard ratio; PD-1, programmed death 1; PD-L1, programmed death ligand 1; SCC, squamous cell carcinoma; TIL, tumor infiltrating lympohocyte.





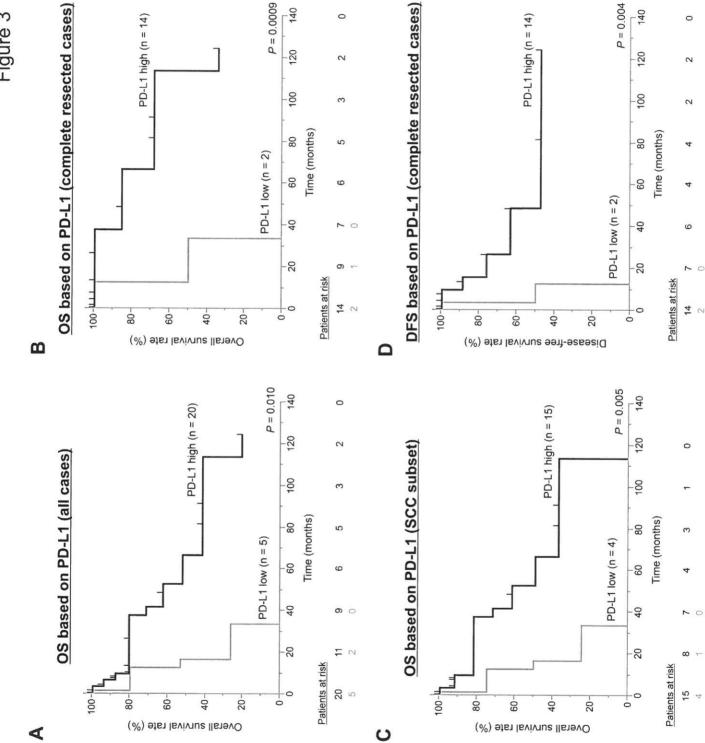
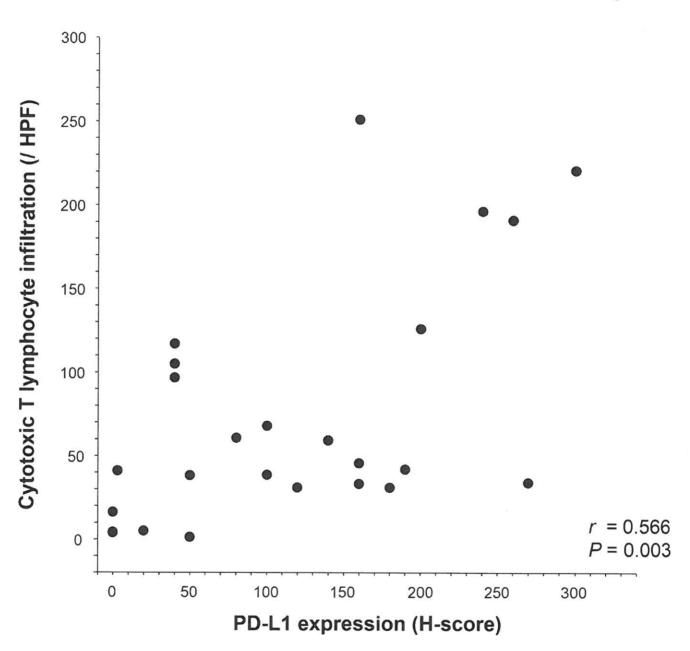


Figure 4



Supplementary Table S1. Patient characteristics

Characteristic	n =	= 25 (%)	
Age, years old			
Median [range]	63	[32-81]	
Sex			
Male	17	68%	
Female	8	32%	
Masaoka-Koga stage			
I	4	16%	
IIa	2	8%	
IIb	1	4%	
III	9	36%	
IVa	3	12%	
IVb	6	24%	
Histological type	<i>3</i> 79		
Squamous cell carcinoma	19	76%	
Adenocarcinoma	4	16%	
Papillary adenocarcinoma	1	4%	
Thymic carcinoma with adenoid cystic carcinoma-like features	1	4%	
Mucinous adenocarcinoma	1	4%	
Adenocarcinoma, NOS	1	4%	
Small cell carcinoma	1	4%	
Sarcomatoid carcinoma	1	4%	
Tumor size, mm		170	
Median [range]	56	[13-90]	
≤ 56 mm	13	52%	
> 56 mm	12	48%	
Curability	122	1070	
Complete resection	16	64%	
Incomplete resection (biopsy)	9	36%	
Neoadjuvant therapy (complete resection cases)		3070	
Chemotherapy	0	0%	
Radiotherapy	2	13%	
Chemoradiotherapy	2	13%	
None	12	75%	
Adjuvant therapy (complete resection cases)	12	1370	
Chemotherapy	1	6%	
Radiotherapy	4	25%	
Chemoradiotherapy	0	0%	
None	11	69%	
Freatments after recurrence (complete resection cases)	11	0976	
Chemotherapy	3	50%	
Radiotherapy	1		
Chemoradiotherapy	0	17% 0%	
Best supportive care	2		
Freatments (incomplete resection cases)	2	33%	
Chemotherapy	2	220/	
Radiotherapy	3	33%	
Chemoradiotherapy	0	0%	
	4	44%	
Best supportive care	2	22%	

Abbreviation: NOS, not otherwise specified.

Supplementary Table S2. Multivariate analysis for prognostic factors affecting overall survival

Characteristic, risk factor	All cases (n = 25)			SCC (n = 19)		
	HR	[95% CI]	P	HR	[95% CI]	P
Masaoka-Koga stage, IVa/IVb	7.000	[1.805-34.979]	0.005	7.000	[1.304-24.008]	0.010
Curability, incomplete resection	7.000	[1.805-34.979]	0.005	7.000	[1.304-24.008]	0.010
PD-L1 expression, low expression	6.854	[1.236-54.411]	0.028	17.091	[1.538-59.367]	0.006
PD-1 positive TILs, per increase by 10 cells / HPF	1.546	[1.063-2.293]	0.026	1.607	[1.077-2.552]	0.023

Abbreviations: CI, confidence interval; HPF, high power field; HR, hazard ratio; PD-1, programmed death 1; PD-L1, programmed death ligand 1; SCC, squamous cell carcinoma; TIL, tumor infiltrating lymphocyte.